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Synthetic Studies on Tumor-Associated Antigens: Efficient Syntheses of Le^a and Sialyl-Le^a Oligosaccharides, and Their Deaminated Analogs¹ Yataka Makimura^a; Hideharu Ishida^a; Makoto Kiso^a; Akira Hasegawa^a ^a Department of Applied Bioorganic Chemistry, Gifu University, Japan

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SYNTHETIC STUDIES ON TUMOR-ASSOCIATED ANTIGENS: EFFICIENT SYNTHESES OF Le^a AND SIALYL-Le^a OLIGOSACCHARIDES, AND THEIR DEAMINATED ANALOGS¹

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ABSTRACT

Efficient syntheses of tumor-associated Le^a, sialyl Le^a and their deaminated analogs are described. The suitably protected D-glucosamine (4) or D-glucose (5) derivative was successively coupled with the methyl-1-thioglycosides (glycosyl donors) of D-galactose (6) and L-fucose (11) in high yields by using Niodosuccinimide/trifluoromethanesulfonic acid (NIS/TfOH) as the glycosyl promoter. The resulting trisaccharides (12 and 13) were each converted, by deprotections, to the Le^a determinant (17) and its deaminated analog (18), and by further glycosylation with the phenyl-2-thioglycoside of N-acetylneuraminic acid (25), to the sialyl-Le^a determinant (30) and its deaminated analog (31), respectively.

INTRODUCTION

The prominent tumor-associated carbohydrate antigens sially Lewis x (sLe^X) and sially Lewis a (sLe^a) have been identified² as the minimal carbohydrate ligands for selectins, a family of lectin-type cell adhesion molecules (LEC-CAMs) involved in

leukocyte trafficking, thrombosis, inflammation, tumor metastasis, and so on. The Lewis a (Le^a) determinant was also detected on human adenocarcinoma cells,³ and the 3'-O-sulfated Le^a structure has recently been suggested⁴ to be a superior oligosaccharide ligand for human E-selectin.

We have succeeded⁵ in a systematic synthesis of sLe^x gangliosides and their analogs by use of a highly efficient, regio- and α -stereoselective sialylation procedure⁶ using 2-thioglycosides of sialic acids in acetonitrile. A number of studies for selectin ligands/inhibitors have been achieved^{2,7} by employing those synthetic sLe^x compounds. Along this line, we have previously reported a total synthesis of sLe^a ganglioside⁸ and some novel epitope analogs containing *N*-methyl-1-deoxynojirimycin.⁹ As a part of our continuing efforts, on the synthesis and elucidation of the biological functions of sialoglycoconjugates, we describe here the efficient syntheses of Le^a (17) and sLe^a (30) oligosaccharides, and their deaminated analogs (18 and 31) by use of thioglycosides as glycosyl donors.

RESULTS AND DISCUSSION

Thioglycosides are highly stable in many organic operations, and capable of specific activations by appropriate thiophilic promoters in very mild conditions, being widely used in oligosaccharide synthesis.¹⁰ In this study, three thioglycosides for D-galactose (6), L-fucose (11) and N-acetylneuraminic acid (25) were successfully employed as efficient glycosyl donors.

Benzyl 4,6-O-benzylidene-2-deoxy-2-phthalimido- β -D-glucopyranoside¹¹ (4) and the corresponding D-glucopyranose derivative¹² (5), which was prepared by the regioselective 3-O-benzoylation of 1 and subsequent 2-O-benzylation, were each coupled with methyl 2,3,4,6-tetra-O-benzoyl-1-thio- β -D-galactopyranoside (6), promoted by N-iodosuccinimide/trifluoromethanesulfonic acid¹³ (NIS/TfOH) in dichloromethane, to give disaccharides 7 (88%) and 9 (94%), respectively (Scheme 1). Reductive ring opening of the benzylidene group in 7 and 9, and subsequent α stereoselective L-fucosylation of 8 and 10 with methyl 2,3,4-tri-O-benzyl-1-thio- β -L-





Scheme 1. i) BzCl, pyr.-CH₂Cl₂, -50 °C; ii) BnBr, Ag₂O, CH₂Cl₂, rt (100%);
iii) NaOMe, MeOH-THF, rt; iv) NIS, TfOH, CH₂Cl₂, -35 °C, 4→7 (88%); 5→9 (94%); v) NaBH₃CN, HCl/Et₂O, THF; vi) NIS, TfOH, CH₂Cl₂, -35 °C, 8→12 (92%), or NIS, TfOH, toluene, -10 °C, 10→13 (90%)

OBn

12 13 R NPhth

OBn

BnÒ



Scheme 2. i) NaOMe, MeOH-CH₂Cl₂ or MeOH-THF; ii) NH₂NH₂•H₂O, 80% EtOH, reflux; iii) Ac₂O, MeOH; iv) H₂, Pd-C, EtOH-AcOH; v) Me₂C(OMe)₂, p-TsOH or CSA, CH₃CN, rt; vi) NaH, BnBr, DMF; vii) 80% AcOH, 45 °C, or p-TsOH, MeOH

fucopyranoside (11) afforded the protected Le^a trisaccharide 12 (92%) and its analog 13 (90%) in high yields, respectively. These were converted, by de-O-benzoylation and hydrogenolytic removal of the benzyl groups, to the desired Le^a determinant 17^{14} and its deaminated analog 18, almost quantitatively (Scheme 2).

Treatment of 15 and 16 with 2,2-dimethoxypropane and a catalytic amount of *p*-toluenesulfonic acid or camphorsulfonic acid in acetonitrile gave 19 (67%) and



Scheme 3. i) NIS, TfOH, CH₃CN, -35 [•]C, 23→26 (52%), 24→27 (59%); ii) NaOMe, MeOH; iii) 0.2M KOH; iv) H₂, Pd(OH)₂-C, 50% aq THF



Scheme 4. i) BzCl, pyr.-CH₂Cl₂, -45 °C; ii) 80% AcOH; iii) NIS, TfOH, CH₃CN, -35 °C (69%); iv) (a) NaOMe, MeOH; (b) 0.2M KOH; (c) H₂, Pd(OH)₂-C, 50% aq THF

21(63%) together with the corresponding 4,6-*O*-isopropylidene derivatives (~30%). Benzylation of the remaining hydroxyls and subsequent removal of the isopropylidene group afforded 23 and 24, which were then coupled with methyl (phenyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-2-thio-D-glycero- α -D-galacto-2-nonulopyranosid) onate¹⁵ (25) in the presence of NIS/TfOH in acetonitrile to give 26 (52%) and 27 (59%), respectively. These were converted stepwise, by de-*O*-acetylation, saponification of the methyl ester and hydrogenolysis over palladium hydroxide on carbon in 50% aq THF, into the desired sLe^a tetrasaccharide 30^{16} and its deaminated analog 31 in almost quantitative yields, respectively (Scheme 3).

We have developed⁶ a facile, and highly regio- and α -stereoselective sialyl glycoside syntheses by using the 2-thioglycosides of sialic acids as glycosyl donors in acetonitrile. In this procedure, the use of lightly protected sugar acceptors, in which several OH groups are unprotected, was another prominent feature.^{6,15b} Treatment of **19** with a restricted amount of benzoyl chloride in pyridine-dichloromethane mixture at -50 °C gave **32**, which was then converted to the desired trisaccharide acceptor **35** (Scheme 4). Iodonium promoted, regio- and α -stereoselective sialylation of **35** with **25** was performed under reaction conditions similar to these employed for **23** to provide the desired tetrasaccharide **3** in about 70% yield. Compound **36** thus obtained was readily converted into **30** as described for **26**.

Until recently, a variety of synthetic studies on carbohydrate probes which contain the Le^a or sLe^a epitope derivatives have been reported.^{15c,17} We here achieved an efficient synthesis of both Le^a and sLe^a oligosaccharides, and their deaminated analogs by employing three thioglycosides 6, 11, and 25 as glycosyl donors. These synthetic oligosaccharides might be useful for further investigation of biological functions.

EXPERIMENTAL

General methods. Optical rotations were determined with a Union PM-201 polarimeter at 25 °C. ¹H NMR spectra were recorded on JEOL JNM-GX 270 (270 MHz) or JNM-GX 400 (400 MHz) spectrometer using deuterated solvents (CDCl3, CD3OD) with TMS as the internal standard. Fast atom bombardment-mass (FAB-MS) spectra were recorded on JEOL JMS-SX 102A mass spectrometer/JMA-DA 7000 data system. Each sample was mixed with triethanolamine (TEA) matrix on a target. The ion accelerating voltage was 8.0 kV, and the primary beam for the bombardment was 6.0 keV of xenon.

All reactions were monitored by TLC (Merck silica gel aluminum plates 60F-254) and preparative column chromatography was performed on silica gel (Wako Chemical Co., 200 mesh) with the solvent systems specified. Concentrations were conducted *in vacuo*.

Benzyl 2-O-Benzyl-4,6-O-benzylidene-β-D-glucopyranoside (5). To a stirred solution of 1 (500 mg, 1.39 mmol) in CH₂Cl₂ (5 mL) and pyridine (0.6 mL) was added benzoyl chloride (1.64 mmol) in CH₂Cl₂ (2 mL) dropwise at -50 °C, and the stirring was continued for 30 min at -50 °C. Methanol was added and the mixture was concentrated. The residue was taken up in CH₂Cl₂, washed with ice-cold M HCl, M Na₂CO₃ and water, dried (Na₂SO₄), and the solvent was evaporated. Column chromatography (1:6 AcOEt-hexane) of the residue on silica gel gave the 3-O-benzoyl derivative 2 (390 mg, 60%); ¹H NMR (CDCl₃) δ 4.64 (d, 1H, J_{1,2} = 7.7 Hz, H-1), 5.48 (t, 1H, J_{2,3} = J_{3,4} = 9.5 Hz, H-3). The 2-O-acetate of **2** showed δ 4.73 (d, 1H, J_{1,2} = 7.9 Hz, H-1), 5.27 (dd, 1H, J_{1,2} = 7.9, J_{2,3} = 9.3 Hz, H-2), 5.56 (t, 1H, J_{2,3} = J_{3,4} = 9.5 Hz, H-3).

To a solution of 2 (4.9 g, 10.6 mmol) in CH₂Cl₂ (50 mL) were added benzyl bromide (3.8 mL, 32 mmol) and silver (I) oxide (14.45 g, 62.4 mmol), and the mixture was stirred in the dark for 10 h at room temperature. Methanol was added and the solids were filtered off through Celite. The filtrate was washed with water, dried, and concentrated to a residue which was chromatographed (1:6 AcOEt-hexane) on a column of silica gel to give **3** (5.85 g, quantitative); ¹H NMR (CDCl₃) δ 3.65 (dd, 1H, J_{1,2} = 7.7, J_{2,3} = 9.2 Hz, H-2), 4.78 (d, 1H, J_{1,2} = 7.7 Hz, H-1), 5.59 (~t, 1H, J_{2,3} = 9.2 Hz, J_{3,4} = 9.5 Hz, H-3). Treatment of **3** (5.85 g) with a catalytic amount of NaOMe in MeOH-THF and column chromatography (CH₂Cl₂) on silica gel afforded the title compound **5** (4.74 g, quantitative): [α]D -42° (*c* 0.8, CHCl₃); ¹H NMR (CDCl₃) δ 3.41 (dd, 1H, J_{1,2} = 7.9, J_{2,3} = 9.2 Hz, H-2), 3.56 (t, 1H, J_{2,3} = J_{3,4} = 9.2 Hz, H-3), 4.63 (d, 1H, J_{1,2} = 7.9 Hz, H-1).

Anal. Calcd for C₂₇H₂₈O₆ (448.5) C, 72.30; H, 6.29. Found: C, 72.52; H, 6.26.

Benzyl O-(2,3,4,6-Tetra-O-benzoyl-β-D-galactopyranosyl)-(1-+3)-4,6-O-benzylidene-2-deoxy-2-phthalimido- β -D-glucopyranoside (7). To a solution of 4 (500 mg, 1.0 mmol) and 6 (1 g, 1.6 mmol) in dry CH₂Cl₂ (5 mL) was added powdered molecular sieves 4Å (MS-4Å, 0.5 g), and the mixture was stirred for 5 h at room temperature, then it was cooled to -35 °C. N-Iodosuccinimide (NIS: 0.67 g, 3 mmol) and trifluoromethanesulfonic acid (TfOH; 53 µL, 0.6 mmol) were added to the cooled mixture, and it was stirred for 3 h at -35 °C and then neutralized with Et3N. The precipitate was filtered off and washed with CH2Cl2. The filtrate and washings were combined, and the solution was successively washed with M Na2S2O3 and water, dried (Na₂SO₄), and concentrated to a syrup which was chromatographed on a column of silica gel (75 g) with 1:3 AcOEt-hexane to give 7 (960 mg, 88%) as an amorphous mass: [a]D +49° (c 0.3, CHCl3); ¹H NMR (CDCl3) & 4.92 (d, 1H, J_{1.2} = 8.1 Hz, H-1b), 5.15 (d, 1H, J_{1,2} = 8.4 Hz, H-1a), 5.35 (dd, 1H, J_{2,3} = 10.3, $J_{3,4} = 3.3 \text{ Hz}, \text{H-3b}$, 5.54 (dd, 1H, $J_{1,2} = 8.1, J_{2,3} = 10.3 \text{ Hz}, \text{H-2b}$), 5.63 (s, 1H, PhCH), 5.80 (near d, 1H, $J_{3,4} = 3.3$ Hz, H-4b), and 6.89-8.05 (m, 34H, aromatic).

Anal. Calcd for C₆₂H₅₁NO₁₆ (1066.1): C, 69.85; H, 4.82; N, 1.31. Found: C, 69.83; H, 4.61; N, 1.15.

Benzyl O-(2,3,4,6-Tetra-O-benzoyl- β -D-galactopyranosyl)-(1-3)-6-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranoside (8). To a solution of 7 (1 g, 0.9 mmol) in dry THF (10 mL) was added MS-4Å (1 g), and the mixture was stirred for 3 h at room temperature, and sodium cyanoborohydride (1 g, 15 mmol) was gradually added. After the reagent had dissolved, hydrogen chloride in ether was added dropwise at 0 °C until the evolution of gas ceased. TLC indicated that the reaction was complete after 15 min and then neutralized with Et₃N. The precipitate was filtered off and washed with CH₂Cl₂ and water. The filtrate and washings were combined, and the solution was washed with water, dried (Na₂SO₄), and concentrated to a syrup which was chromatographed on a column of silica gel (75 g) with 1:2 AcOEt-hexane to give 8 (900 mg, 90%) as an amorphous mass: [α]D +54° (c 0.16, CHCl₃); ¹H NMR (CDCl₃) δ 3.47 (bd, 1H, OH-4a), 4.82 (d, 1H, J_{1,2} = 8.1 Hz, H- 1b), 4.95 (d, 1H, $J_{1,2} = 8.4$ Hz, H-1a), 5.47 (dd, 1H, $J_{2,3} = 10.3$, $J_{3,4} = 3.3$ Hz, H-3b), 5.74 (dd, 1H, H-2b), 5.88 (d, 1H, H-4b), and 6.85-8.15 (m, 34H, aromatic).

Anal. Calcd for C₆₂H₅₃NO₁₆ (1068.1): C, 69.72; H, 5.00; N, 1.31. Found: C, 69.59; H, 4.98; N, 1.06.

Benzyl *O*-(2,3,4,6-Tetra-*O*-benzoyl-β-D-galactopyranosyl)-(1→3)-2-*O*-benzyl-4,6-*O*-benzylidene-β-D-glucopyranoside (9). To a solution of 5 (1 g, 2.2 mmol) and 6 (2.1 g, 3.3 mmol) in dry CH₂Cl₂ (20 mL) was added MS-4Å (2 g), and the mixture was stirred for 8 h at room temperature, then it was cooled to -35 °C. NIS (1 g, 4.4 mmol) and TfOH (79 µL, 0.89 mmol) were added to the cooled mixture, and it was stirred for 30 min at -35 °C and then neutralized with Et₃N. A workup similar to that described for 7 gave 9 (2.16 g, 94%) as an amorphous mass: $[\alpha]_D$ +9° (*c* 1.2, CHCl₃); ¹H NMR (CDCl₃) δ 4.57 (d, 1H, J_{1,2} = 7.9 Hz, H-1a), 5.24 (d, 1H, J_{1,2} = 7.9 Hz, H-1b), 5.56 (dd, 1H, J_{2,3} = 10.3 Hz, J_{3,4} = 3.3 Hz, H-3b), 5.61 (s, 1H, PhCH), 5.85 (dd, 1H, H-2b), 5.94 (d, 1H, H-4b) and 7.1-8.1 (m, 35H, 7Ph).

Anal. Calcd for C₆₁H54O₁₅ (1027.0): C, 71.33; H, 5.30. Found: C, 71.17; H, 5.07.

Benzyl O-(2,3,4,6-Tetra-O-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-2,6-di-O-benzyl- β -D-glucopyranoside (10). To a solution of 9 (190 mg, 0.18 mmol) in dry THF (1.9 mL) was added MS-4Å (0.19 g), and the mixture was stirred for 1 h at room temperature, and sodium cyanoborohydride (170 mg, 2.7 mmol) was gradually added. After the reagent had dissolved, hydrogen chloride in ether was added dropwise at 0 °C until the evolution of gas ceased. TLC indicated that the reaction was complete after 5 min and then neutralized with Et3N. A workup similar to that described for 8 gave 10 (160 mg, 84%) as an amorphous mass: [α]D +56° (c 1.0, CHCl3); ¹H NMR (CDCl3) δ 4.43 (d, 1H, J_{1,2} = 7.7 Hz, H-1a), 5.15 (d, 1H, J_{1,2} = 7.9 Hz, H-1b), 5.63 (dd, 1H, J_{2,3} = 10.4, J_{3,4} = 3.5 Hz, H-3b), 5.89 (dd, 1H, H-2b), 5.98 (d, 1H, H-4b), and 7.0-8.1 (m, 35H, 7Ph).

Anal. Calcd for C₆₁H₅₆O₁₅ (1029.1): C, 71.20; H, 5.49. Found: C, 71.17; H, 5.48.

Benzyl *O*-(2,3,4,6-Tetra-*O*-benzoyl-β-D-galactopyranosyl)-(1→3)-*O*-[2,3,4-tri-*O*-benzyl-α-L-fucopyranosyl-(1 → 4)]-6-*O*-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranoside (12). To a solution of 8 (650 mg, 0.63 mmol) and 11 (583 mg, 1.25 mmol) in dry CH₂Cl₂ (6 mL) was added MS-4Å (0.6 g), and the mixture was stirred for 5 h at room temperature. It was cooled to -40 °C. NIS (790 mg, 3.75 mmol) and TfOH (44 μ L, 0.31 mmol) were added to the cooled mixture, and it was stirred for 3 h at -40 °C and then neutralized with Et₃N. A workup similar to that described for 7 gave 12 (860 mg, 92%) as an amorphous mass: [α]D +4° (c 0.7, CHCl₃); ¹H NMR (CDCl₃) δ 1.52 (d, 3H, J₅,6 = 6.6 Hz, H-6c), 4.82 (d, 1H, J_{1,2} = 8.2 Hz, H-1b), 5.18 (dd, 1H, J_{2,3} = 10.4, J₃, 4 = 3.7 Hz, H-3b), 5.25 (bd, 1H, J_{1,2} = ~3 Hz H-1c), 5.69 (dd, 1H, H-2b), 5.78 (d, 1H, H-4b), and 6.85-8.12 (m, 49H, aromatic).

Anal. Calcd for C₈₉H₈₁NO₂₀ (1484.6): C, 72.00; H, 5.50; N, 0.94. Found: C, 71.93; H, 5.40; N, 0.74.

Benzyl O-(2,3,4,6-Tetra-O-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-O-[2,3,4-tri-O-benzyl- α -L-fucopyranosyl-(1 \rightarrow 4)]-2,6-di-O-benzyl- β -Dglucopyranoside (13). To a solution of 10 (500 mg, 0.48 mmol) and 11 (340 mg, 0.72 mmol) in dry toluene (9 mL) was added MS-4Å (0.9 g), and the mixture was stirred overnight at room temperature, then it was cooled to -15 °C. NIS (170 mg, 0.72 mmol) and TfOH (6 μ L, 60 mmol) were added to the cooled mixture, and it was stirred overnight at -10 °C and then neutralized with Et₃N. A workup similar to that described for 12 gave 13 (630 mg, 90%) as an amorphous mass: [α]D -25° (c 0.4, CHCl₃); ¹H NMR (CDCl₃) δ 1.59 (d, 3H, J₅,6 = 6.4 Hz, H-6c), 4.34 (d, 1H, J₁,2 = 7.7 Hz, H-1a), 5.16 (d, 1H, J₁,2 = \sim 3 Hz, H-1c), 5.57 (dd, 1H, J₂,3 = 10.3, J₃,4 = 3.3 Hz, H-3b), 5.57 (d, 1H, J₁,2 = 8.2 Hz, H-1b overlapping with H-3b), 5.85 (dd, 1H, H-2b), 5.89 (d, 1H, H-4b), and 7.1-8.1 (m, 50H, 10Ph).

Anal. Calcd for C₈₈H₈₄O₁₉ (1445.6): C, 73.12; H, 5.86. Found: C, 73.08; H, 5.65.

Benzyl O-(β -D-Galactopyranosyl)-($1 \rightarrow 3$)-O-[2,3,4-tri-O-benzyl- α -L-fucopyranosyl-($1 \rightarrow 4$)]-6-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranoside (14). To a solution of 12 (230 mg, 0.16 mmol) in MeOH (4 mL) and CH₂Cl₂ (1 mL) was added catalytic amount of sodium methoxide, and the mixture was stirred for 5 h at room temperature, and then neutralized with Amberlite IR-120 (H⁺) resin and filtered. The resin was washed with MeOH and the combined filtrate and washings were concentrated. The residue was chromatographed on a column of silica gel (30 g) with 25:1 CH₂Cl₂-MeOH to give 14 (163 mg, quantitative) as an amorphous mass: $[\alpha]_D$ -63° (c 0.4, CHCl₃); ¹H NMR (CDCl₃) δ 1.61 (d, 3H, J_{5,6} = 6.6 Hz, H-6c), 5.22 (d, 1H, J_{1.2} = 3.3 Hz, H-1c), 7.22-7.78 (m, 29H, aromatic).

Anal. Calcd for C₆₁H₆₅NO₁₆ (1068.2): C,68.59; H, 6.13; N, 1.31. Found: C,68.48; H, 5.92; N, 1.30.

Benzyl $O-(\beta-D-Galactopyranosyl)-(1\rightarrow 3)-O-[2,3,4-tri-O-benzyl-\alpha L-fucopyranosyl-(1 <math>\rightarrow$ 4)]-2-acetamido-6-O-benzyl-2-deoxy- β -D-glucopyranoside (15). To a solution of 14 (360 mg, 0.34 mmol) in aq 80% EtOH (4 mL) was added hydrazine hydrate (0.34 mL), the mixture was refluxed for 5 h. After cooling the mixture in ice bath, the precipitate was filtered off, and washed with MeOH. The filtrate and washings were combined, and concentrated. The residue thus obtained was then treated with acetic anhydride (0.16 mL) in MeOH (3 mL) for 4 h at room temperature. The reaction mixture was then concentrated to a syrup which was chromatographed on a column of silica gel (30 g) with 25:1 CH₂Cl₂-MeOH, to give 15 (330 mg, quantitative) as an amorphous mass: $[\alpha]_D$ -67° (c 1.5, CHCl₃); ¹H NMR (CDCl₃) δ 1.22 (d, 3H, J_{5,6} = 6.4 Hz, H-6c), 2.00 (s, 3H, AcN), 5.25 (d, 1H, J_{1,2} = 3.5 Hz, H-1c), and 7.23-7.31 (m, 25H, 5Ph).

Anal. Calcd for C55H65NO15 (980.1): C, 67.47; H, 6.68; N, 1.43. Found: C, 67.27; H, 6.41; N, 1.15.

Benzyl O-(β -D-Galactopyranosyl)-($1\rightarrow 3$)-O-[2,3,4-tri-O-benzyl- α -L-fucopyranosyl-($1\rightarrow 4$)]-2,6-di-O-benzyl- β -D-glucopyranoside (16). To a solution of 13 (1.3 g, 0.9 mmol) in MeOH (13 mL) and THF (6 mL) was added catalytic amount of sodium methoxide, and the mixture was stirred for a day at room temperature. It was then neutralized with Amberlite IR-120 (H⁺) resin and filtered. A workup similar to that described for 14 gave 16 (820 mg, 89%) as an amorphous mass: [α]_D -64° (*c* 1.1, CHCl₃); ¹H NMR (CDCl₃) δ 1.12 (d, 3H, H-6c), 5.14 (d, 1H, H-1c), and 7.25-7.77 (m, 30H, 6Ph).

Anal. Calcd for C₆₀H₆₈O₁₅ (1029.2): C, 70.02; H, 6.66. Found: C, 69.95; H, 6.45.

O-(β-D-Galactopyranosyl)-(1→3)-*O*-[α-L-fucopyranosyl-(1→4)]-2acetamido-2-deoxy-D-glucopyranose (17). Compound 15 (50 mg, 0.05 mmol) in EtOH (1.3 mL) and AcOH (0.2 mL) was hydrogenolyzed in the presence of 10% Pd-C (50 mg) for a day at 45 °C, then the mixture was filtered, and concentrated. Chromatography (MeOH) of the residue on Sephadex LH-20 (80 g) gave 17 (21 mg, 78%) as an amorphous mass: [α]D -32° (*c* 0.5, MeOH) [ref. 14, -45.1° (*c* 1, water)]; ¹H NMR (CD3OD) δ 1.19 (d, 3H, J5,6 = 6.6 Hz, H-6c), 1.99 (s, 3H, AcN), 4.39 (d, 0.19H, J_{1,2} = 7.3 Hz, H-1b of β-anomer), 4.45 (d, 0.81H, J_{1,2} = 7.3 Hz, H-1b of α-anomer), 4.64 (d, 0.19H, J_{1,2} = 8.1 Hz, H-1a of β-anomer), 4.89 (q, 1H, H-5c), 4.99 (d, 1H, J_{1,2} = 2.6 Hz, H-1c) and 5.04 (d, 0.81H, J_{1,2} = 4.0 Hz, H-1a of α-anomer). FAB-MS (negative ion mode): *m/z* 528.2 (M-H)⁻. Average mass of M-H was 528.4877 for C₂₀H₃₄NO₁₅.

Anal. Calcd for C₂₀H₃₅NO₁₅ (529.5): C, 45.37; H, 6.66; N, 2.65. Found: C, 45.11; H, 6.58; N, 2.61.

O-(β-D-Galactopyranosyl)-(1→3)-*O*-[α-L-fucopyranosyl-(1→4)]-β-D-glucopyranose (18). Compound 16 (100 mg, 0.1 mmol) in EtOH (2.5 mL) was hydrogenolyzed in the presence of 20% Pd(OH)₂-C (100 mg) for a day at room temperature. A workup similar to that described for 17 gave 18 (41 mg, 86%) as an amorphous mass: $[\alpha]_D$ -39° (*c* 0.8, MeOH); ¹H NMR (CD₃OD) δ 1.18 (d, 3H, J_{5,6} = 6.59 Hz, H-6c), 4.51 (d, 0.55H, J_{1,2} = 7.9 Hz, H-1b of β-anomer), 4.70 (d, 0.45H, J_{1,2} = 7.7 Hz, H-1b of α-anomer), 4.74 (d, 0.55H, J_{1,2} = 7.7 Hz, H-1a of β-anomer), 4.89 (q, 1H, J = 6.6 Hz, H-5c), 5.02 (d, 1H, J_{1,2} = 3.7 Hz, H-1c), and 5.11 (d, 0.45H, J_{1,2} = 3.7 Hz, H-1a of α-anomer). FAB-MS (negative ion mode): m/z 487.2 (M-H)⁻. Average mass of M-H was 487.4351 for C18H31O15.

Anal. Calcd for C₁₈H₃₂O₁₅ (488.4): C, 44.26; H, 6.60. Found: C, 44.09; H, 6.31.

Benzyl *O*-(3,4-*O*-Isopropylidene-β-D-galactopyranosyl)-(1-3)-*O*-[2,3,4-tri-*O*-benzyl-α-L-fucopyranosyl-(1-4)]-2-acetamido-6-*O*-benzyl-2-deoxy-β-D-glucopyranoside (19). To a solution of 15 (140 mg, 0.14 mmol) in CH₃CN (1.5 mL) were added 2,2-dimethoxypropane (40 µL) and a catalytic amount of *p*-toluenesulfonic acid (*p*-TsOH). The mixture was stirred for 4 h at room temperature, then neutralized with Amberlite IR-410 (OH⁻) and filtered. The resin was washed with MeOH and the combined filtrate and washings were concentrated. The residue was chromatographed on a column of silica gel (30 g) with 40:1 CH₂Cl₂-MeOH, to give **19** (98 mg, 67%) as an amorphous mass with the corresponding 4,6-*O*-isopropylidene derivative (26%). Compound **19** had [α]_D-62° (*c* 1.3, CHCl₃); ¹H NMR (CD₃OD) δ 1.12 (d, 3H, J5,6 = 6.4 Hz, H-6c), 1.31,1.37 (2s, 6H, Me₂C), 1.91 (s, 3H, AcN), 4.36 (d, 1H, J_{1,2} = 8.2 Hz, H-1b), 5.09 (d, 1H, J_{1,2} = 3.5 Hz, H-1c), and 7.2-7.4 (m, 25H, 5Ph).

Anal. Calcd for C58H69NO15 (1020.2): C, 68.29; H, 6.82; N, 1.37. Found: C, 68.14; H, 6.70; N, 1.25.

Benzyl O-(2,6-Di-O-benzyl-3,4-O-isopropylidene- β -D-galactopyranosyl) - (1 \rightarrow 3) - O-[2,3,4- tri-O-benzyl- α -L-fucopyranosyl- (1 \rightarrow 4)]-2acetamido-6-O-benzyl-2-deoxy- β -D-glucopyranoside (20). To a solution of 19 (63.7 mg, 0.06 mmol) in dry DMF (1 mL), was added sodium hydride (6 mg, 0.15 mmol). After the reagent had dissolved, benzyl bromide was added, and the mixture was stirred for 1 h at 0 °C. The precipitate was filtered off and washed with MeOH. The combined filtrate and washings were concentrated to a syrup which was chromatographed on a column of silica gel (30 g) with 1:2 AcOEt-hexane to give 20 (52 mg, 70%) as an amorphous mass: [α]_D -35° (c 1.0, CHCl₃); ¹H NMR (CDCl₃) δ 1.04 (d, 3H, J_{5,6} = 6.4 Hz, H-6c), 1.29, 1.33, (2s, 6H, Me₂C), 1.62 (s, 3H, AcN), 5.17 (d, 1H, J_{1,2} = 3.7 Hz, H-1c), 6.24 (d, 1H, J_{NH,2} = 7.9 Hz, NH), and 7.15-7.4 (m, 35H, 7Ph).

Anal. Calcd for C₇₂H₈₁NO₁₅ (1200.4): C, 72.04; H, 6.80; N, 1.17. Found: C, 71.96; H, 6.52; N, 1.14.

Benzyl $O.(3,4-O.Isopropylidene-\beta-D-galactopyranosyl)-(1\rightarrow3)-O-$ [2,3,4- tri- O- benzyl- α - L- fucopyranosyl- $(1 \rightarrow 4)$]- 2,6- di-O-benzyl- β -Dglucopyranoside (21). To a solution of 16 (100 mg, 0.1 mmol) in CH₃CN (1 mL) was added 2,2-dimethoxypropane (27 µL) and a catalytic amount of camphorsulfonic acid (CSA), and the mixture was stirred for 2 h at room temperature. A workup similar to that described for 19 gave 21 (65 mg, 63%) as an amorphous mass with the corresponding 4,6-O-isopropylidene derivative (32%). Compound 21 had [α]D -48° (c 1.3, CHCl₃); ¹H NMR (CDCl₃) δ 1.11 (d, 3H, J_{5,6} = 6.6 Hz, H-6c), 1.33,1.42 (2s, 6H, Me₂C), 3.94 (dd, 1H, H-2c), 5.14 (d, 1H, J_{1,2} = 3.5 Hz, H-1c), and 7.24-7.40 (m, 30H, 6Ph).

Anal. Calcd for C₆₃H₇₂O₁₅ (1069.3): C, 70.77; H, 6.79. Found: C, 70.58; H, 6.73.

Benzyl O-(2,6-Di-O-benzyl-3,4-O-isopropylidene- β -D-galactopyranosyl-(1 \rightarrow 3)-O-[2,3,4-tri-O-benzyl- α -L-fucopyranosyl-(1 \rightarrow 4)]-2,6-di-O-benzyl- β -D-glucopyranoside (22). To a solution of 21 (200 mg, 0.2 mmol) in dry DMF (2 mL), was added sodium hydride (22 mg, 0.6 mmol). After the reagent had dissolved, benzyl bromide (0.73 mL) was added, and the mixture was stirred for 1 h at room temperature. A workup similar to that described for 20 gave 22 (220 mg, 94%) as an amorphous mass: [α]D -2° (c 1.2, CHCl₃); ¹H NMR (CDCl₃) δ 1.05 (d, 3H, J₅,6 = 6.6 Hz, H-6c), 1.30,1.35 (2s, 6H, Me₂C), 5.10 (d, 1H, J_{1,2} = 3.5 Hz, H-1c), 7.19-7.35 (m, 40H, 8Ph).

Anal. Calcd for C77H84O15 (1249.5): C, 74.02; H, 6.78. Found: C, 73.82; H, 6.77.

Benzyl O- $(2, 6 - \text{Di} - O - \text{benzyl} - \beta - D - \text{galactopyranosyl}) - (1 \rightarrow 3) - O-$ [2,3,4-tri-O-benzyl- α -L-fucopyranosyl- $(1\rightarrow 4)$]-2-acetamido-6-O-benzyl-2-deoxy- β -D-glucopyranoside (23). A solution of 20 (52 mg, 0.04 mmol) in 80% aq AcOH (1 mL) was heated for 3 h at 45 °C, and concentrated to give 23 (50 mg, quantitative) as an amorphous mass: $[\alpha]_D$ -41° (c 1.1, CHCl3); ¹H NMR (CDCl3) δ 1.10 (d, 3H, J5,6 = 6.41 Hz, H-6c), 1.62 (s, 3H, AcN), 5.21 (d, 1H, J_{1,2} = 3.8 Hz, H-1c), and 7.21-7.34 (m, 35H, 7Ph). Anal. Calcd for C69H77NO15 (1160.4): C, 71.42; H, 6.69; N, 1.21. Found: C, 71.25; H, 6.49; N, 1.07.

Benzyl O- (2,6 - Di - O - benzyl - β - D - galactopyranosyl) - (1 \rightarrow 3) - O-[2,3,4-tri-O-benzyl- α -L-fucopyranosyl-(1 \rightarrow 4)]-2,6-di-O-benzyl- β -Dglucopyranoside (24). To a solution of 22 (447 mg, 0.36 mmol) in MeOH (15 mL) was added catalytic amount of p-TsOH, and the mixture was stirred for 3 h at 45 °C. A workup similar to that described for 19 to give 24 (369 mg, 85%) as an amorphous mass: [α]D -43° (c 0.9, CHCl₃); ¹H NMR (CDCl₃) δ 1.20 (d, 3H, J_{5,6} = 6.59 Hz, H-6c), 4.43 (d, 1H, J_{1,2} = 7.7 Hz, H-1a), 5.04 (d, 1H, J_{1,2} = 7.3 Hz, H-1b), 5.11 (d, 1H, J_{1,2} = 3.5 Hz, H-1c), and 7.17-7.35 (m, 40H, 8Ph).

Anal. Calcd for C74H80O15 (1209.4): C, 73.49; H, 6.67. Found: C,73.25; H, 6.62.

Benzyl O - (Methyl 5 - Acetamido - 4,7,8,9 - tetra - O - acetyl - 3,5 - dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2+3)-O-(2,6-di-O-benzyl- β -D galactopyranosyl) - (1 + 3) - O-[2,3,4- tri-O-benzyl- α -L-fucopyranosyl-(1 + 4)]-2-acetamido-6-O-benzyl-2-deoxy- β -D-glucopyranoside (26). To a solution of 23 (104 mg, 0.09 mmol) and 25 (110 mg, 0.18 mmol) in dry CH₃CN (2 mL) was added MS-4Å (0.2 g), and the mixture was stirred for 8 h at room temperature, then it was cooled to -35 °C. NIS (84 mg, 0.38 mmol) and TfOH (6 μ L, 0.06 mmol) were added to the cooled mixture, and it was stirred for 3 h at -35 °C and then neutralized with Et₃N. A workup similar to that described for 7 gave 26 (76 mg, 52%) as an amorphous mass: [α]D -53° (c 0.72, CHCl₃); ¹H NMR (CDCl₃) δ 1.12 (d, 3H, J₅,6 = 6.4 Hz, H-6c), 1.65, 1.88, 2.00, 2.01, 2.02, 2.10 (6s, 18H, 4AcO, 2AcN), 2.54 (dd, 1H, J_{gem} = 13, J_{3eq},4 = 4.4 Hz, H-3deq), 3.76 (s, 3H, CO₂Me), 5.18 (d, 1H, J_{1,2} = 3.5 Hz, H-1c), 5.34 (dd, 1H, J_{6,7} = 2.4, J_{7,8} = 8.3 Hz, H-7d), 5.42 (m, 1H, H-8d), 5.17, 6.12 (2d, 2H, NH), and 7.23-7.37 (m, 35H, 7Ph).

Anal. Calcd for C89H104N2O27 (1633.8): C, 65.43; H, 6.42; N, 1.71. Found: C, 65.42; H, 6.41; N, 1.56. A sample of 26 (10 mg) was acetylated with acetic anhydride (0.2 mL) and pyridine (0.2 mL) to give the corresponding acetate as an amorphous mass, which showed a narrow doublet (J_{3,4} = 3.0 Hz, H-4b) at δ 5.03 in the ¹H NMR (CDCl₃) spectrum.

Benzyl O - (Methyl 5 - Acetamido - 4,7,8,9 - tetra - O - acetyl - 3,5 dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-O-(2,6di-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-O-[2,3,4-tri-O-benzyl- α -Lfucopyranosyl-(1 \rightarrow 4)]-2,6-di-O-benzyl- β -D-glucopyranoside (27). To a solution of 24 (100 mg, 0.08 mmol) and 25 (104 mg, 0.18 mmol) in dry CH₃CN (2 mL) was added MS-4Å (0.2 g), and the mixture was stirred for 8 h at room temperature, then it was cooled to -35 °C. NIS (80 mg, 0.36 mmol) and TfOH (6 μ L, 0.14 mmol) were added to the cooled mixture, and the mixture was stirred for 4 h at -35 °C and then neutralized with Et₃N. A workup similar to that described for 7 gave 27 (82 mg, 59%) as an amorphous mass: [α]D -32° (c 1.6, CHCl₃); ¹H NMR (CDCl₃) δ 1.21 (d, 3H, J5,6 = 6.4 Hz, H-6d), 1.90, 1.92, 1.96, 2.00, 2.10 (5s, 15H, 4AcO, AcN), 2.49 (dd, 1H, J_{gem} = 13, J_{3eq,4} = 4.5 Hz, H-3deq), 3.82 (s, 3H, CO₂Me), 4.46 (d, 1H, J_{1,2} = 8.3 Hz, H-1a), 5.04 (d, 1H, J_{1,2} = 7.9 Hz, H-1b), 5.08 (d, 1H, J_{1,2} = 2.4 Hz, H-1c), and 7.15-7.40 (m, 40H, 8Ph).

Anal. Calcd for C94H107NO27 (1682.9): C, 67.09; H, 6.41; N, 0.83. Found: C, 66.92; H, 6.24; N, 0.69.

Benzyl $O \cdot (5 \cdot \text{Acetamido} - 3, 5 \cdot \text{dideoxy} - D \cdot glycero \cdot \alpha \cdot D \cdot galacto - 2 \cdot \text{nonulopyranosylonic acid} \cdot (2 \rightarrow 3) \cdot O \cdot (2, 6 \cdot \text{di} \cdot O \cdot \text{benzyl} - \beta - D \cdot \text{galacto-pyranosyl} - (1 \rightarrow 3) \cdot O \cdot [2, 3, 4 \cdot \text{tri} \cdot O \cdot \text{benzyl} - \alpha - \text{L-fucopyranosyl} - (1 \rightarrow 4)] - 2 \cdot \text{acetamido} - 6 \cdot O \cdot \text{benzyl} - 2 \cdot \text{deoxy} - \beta - D \cdot \text{glucopyranoside} (28).$ To a solution of 26 (37 mg, 0.02 mmol) in MeOH (2 mL) was added catalytic amount of sodium methoxide, and the mixture was stirred for a day at room temperature. 0.2M-KOH (0.5 mL) was added to the mixture and stirring was continued for 6 h at room temperature. The solution was neutralized with Amberlite IR-120 (H⁺) resin and filtered. The resin was washed with MeOH and the combined filtrate and washings were concentrated. The residue was chromatographed on a column of silica gel (30 g)

with 20:1 CH₂Cl₂-MeOH, to give **28** (30 mg, 91%) as an amorphous mass: $[\alpha]_D$ -61° (*c* 0.4, CHCl₃); ¹H NMR (CDCl₃) δ 1.01 (d, 3H, J_{5,6} = 6.4 Hz, H-6c), 1.86, 1.98 (2s, 6H, 2AcN), 2.80 (dd, 1H, J_{gem} = 13, J_{3eq,4} = 4 Hz, H-3deq), 5.01 (d, 1H, J_{1,2} = 3.8 Hz, H-1c), and 7.10-7.29 (m, 35H, 7Ph).

Anal. Calcd for C₈₀H94N₂O₂₃ (1451.6): C, 66.19; H, 6.53; N, 1.93. Found: C, 65.90 H, 6.41; N, 1.74.

Benzyl O-(5 - Acetamido - 3,5 - dideoxy - D - glycero - α - D - galacto-2-nonulopyranosylonic acid) - (2 \rightarrow 3) - O-(2,6- di-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-O-[2,3,4-tri-O-benzyl- α -L-fucopyranosyl-(1 \rightarrow 4)]-2,6-di-O-benzyl- β -D-glucopyranoside (29). To a solution of 27 (86 mg, 0.05 mmol) in MeOH (2 mL) was added a catalytic amount of sodium methoxide, and the mixture was stirred for a day at room temperature. 0.2M KOH (0.5 mL) was added to the mixture, and the stirring was continued for a day at room temperature. The solution was neutralized with Amberlite IR-120 (H⁺) resin and filtered. A workup similar to that described for 28 gave 29 (60 mg, 78%) as an amorphous mass: [α]D -31° (c 1.20, CHCl₃); ¹H NMR (CD₃OD) δ 1.23 (d, 3H, J₅,6 = 6.4 Hz, H-6c), 1.94 (t, 1H, Jgem = J_{3ax},4 = 13 Hz, H-3dax), 2.04 (s, 3H, AcN), 2.92 (dd, 1H, Jgem = 13, J_{3eq},4 = 4.5 Hz, H-3deq), 4.35 (d, 1H, J₁,2 = 7.9 Hz, H-1a), 4.96 (d, 1H, J₁,2 = 7.9 Hz, H-1b), 5.02 (d, 1H, J₁,2 = 3.5 Hz, H-1c), and 7.15-7.48 (m, 40H, 8Ph).

Anal. Calcd for C₈₅H97NO₂₃ (1500.7): C, 68.03; H, 6.52; N, 0.93. Found: C, 67.78; H, 6.39; N, 0.92.

O-(5-Acetamido-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonic acid)-(2→3)-*O*-(β-D-galactopyranosyl)-(1→3)-*O*-[α-L-fucopyranosyl-(1→4)]-2-acetamido-2-deoxy-D-glucopyranose (30). Compound 28 (37 mg, 0.03 mmol) in THF (1 mL) and H₂O (1 mL) was hydrogenolyzed in the presence of 20% Pd(OH)₂-C (10 mg) for a day at room temperature, then filtered, and concentrated. Chromatography (MeOH) of the residue on Sephadex LH-20 (80 g) gave 30 (23 mg, quantitative) as an amorphous mass: $[\alpha]_D$ -33° (*c* 0.8, CH₃OH); ¹H NMR (CD₃OD) δ 1.18 (d, 3H, J_{5,6} = 6.4 Hz, H-6c), 1.98, 2.03 (2s, 6H, 2AcN), 2.82 (dd, 1H, J_{gem} = 12, J_{3eg}, 4 = 4.4 Hz, H-3deq), 4.39 (d, 0.27H, J_{1,2} = 7.5 Hz, H-1b of β-anomer), 4.45 (d, 0.73H, $J_{1,2} = 7.2$ Hz, H-1b of α-anomer), 4.63 (d, 0.27H, $J_{1,2} = 7.9$ Hz, H-1a of β-anomer), 4.91 (q, 1H, H-5c), 4.99 (d, 0.73H, $J_{1,2} = 3.1$ Hz, H-1c of α-anomer) and 5.03 (d, 0.73H, $J_{1,2} = 4$ Hz, H-1a of α-anomer). FAB-MS (negative ion mode): m/z 819.3 (M-H)⁻. Average mass of M-H was 819.7455 for C₃₁H₅₁N₂O₂₃.

Anal. Calcd for C₃₁H₅₂N₂O₂₃ (820.8): C, 45.37; H, 6.39; N, 3.41. Found: C, 45.12; H, 6.15; N, 3.22.

O-(5-Acetamido-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonic acid)-(2→3)-*O*-(β-D-galactopyranosyl)-(1→3)-*O*-[α-L-fucopyranosyl-(1→4)]-β-D-glucopyranose (31). Compound 29 (58 mg, 0.04 mmol) in THF (1 mL) and H₂O (1 mL) was hydrogenolyzed in the presence of 20% Pd(OH)₂-C (10 mg) for a day at room temperature, and the mixture was filtered. A workup similar to that described for 30 gave 31 (28 mg, 93%) as an amorphous mass: [α]_D-17° (*c* 0.6, MeOH); ¹H NMR (CD₃OD) δ 1.16 (d, 3H, J₅,6 = 6.2 Hz, H-6c), 2.00 (s, 3H, AcN), 2.85 (dd, 1H, Jgem = 13, J_{3eq,4} = 4 Hz, H-3deq), 4.50 (d, 0.56H, J_{1,2} = 7.7 Hz, H-1b of β-anomer), 4.73 (d, 0.44H, J_{1,2} = 7.7 Hz, H-1b of α-anomer), 4.77 (d, 0.56H, J_{1,2} = 7.7 Hz, H-1a of β-anomer), 5.01 (~t, 1H, J_{1,2} = 3~4 Hz, H-1c of two anomers are overlapping), and 5.11 (d, 0.44H, J_{1,2} = 3.5 Hz, H-1a of α-anomer). FAB-MS (negative ion mode): *m/z* 778.3 (M-H)⁻. Average mass of M-H was 778.6930 for C29H48NO23.

Anal. Calcd for C₂₉H₄₉NO₂₃ (779.7): C, 44.67; H, 6.33; N, 1.80. Found: C, 44.64; H, 6.30; N, 1.50.

Benzyl $O \cdot (6 \cdot O \cdot Benzoyl \cdot 3, 4 \cdot O \cdot isopropylidene \cdot \beta \cdot D \cdot galactopyrano$ $syl) - <math>(1 \rightarrow 3) \cdot O \cdot [2,3,4 \cdot tri \cdot O \cdot benzyl \cdot \alpha \cdot L \cdot fucopyranosyl \cdot <math>(1 \rightarrow 4)] \cdot 2 \cdot$ acetamido - 6 - O - benzyl - 2 - deoxy - \beta - D - glucopyranoside (32). To a stirred solution of 19 (140 mg, 0.14 mmol) in CH₂Cl₂ (1.4 mL) and pyridine (0.1 mL) was added benzoyl chloride (0.21 mmol) in CH₂Cl₂ (0.5 mL) dropwise at -50 °C, and the stirring was continued for 1 h at -50 °C. A workup similar to that described for 5 and column chromatography (100:1 CH₂Cl₂-MeOH) on silica gel gave 32 (80 mg, 52%) together with 33 (8%) and 34 (38%). Compound 32 had [α]D -53° (c 1.7, CHCl₃); ¹H NMR (CDCl₃) δ 1.10 (d, 3H, J_{5,6} = 6.2 Hz, H-6c), 1.35, 1.52, 1.53 (3S, 9H, Me₂C and AcN), 5.14 (d, 1H, J_{1,2} = 3.7 Hz, H-1c), 6.68 (d, 1H, J_{NH,2} = 8.3 Hz, NH), 7.20-7.45 (m, 25H of 5Bn and 3H of Bz), and 8.00 (d, 2H, J = 7.33, *o*-position of Bz).

Anal. Calcd for C65H73NO16 (1124.3): C, 69.44; H, 6.54; N, 1.25. Found: C, 69.35; H, 6.26; N, 1.11.

Benzyl O-(6-O-Benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-O-[2,3,4-tri-O-benzyl- α -L-fucopyranosyl-(1 \rightarrow 4)]-2-acetamido-6-O-benzyl-2-deoxy- β -D-glucopyranoside (35). A mixture of 32 (96.5 mg, 0.088 mmol) and 80% aq AcOH (1 mL) was stirred for a day at 45 °C. The solvent was removed by evaporation (with toluene) to give 35 (91.3 mg, quantitative) as an amorphous mass: [α]_D -55° (*c* 1.6, CHCl₃); ¹H NMR (CDCl₃) δ 1.12 (d, 3H, J_{5,6} = 6.2 Hz, H-6c), 1.58 (S, 3H, AcN), 5.12 (d, 1H, J_{1,2} = 3.7 Hz, H-1c), 6.75 (d, 1H, J_{NH,2} = 8.3 Hz, NH), 7.20-7.45 (m, 25H of 5Bn and 3H of Bz), and 7.99 (d, 2H, J = 7.2 Hz, *o*-position of Bz).

Anal. Calcd for C₆₂H₆₉NO₁₆ (1084.2): C, 68.68; H, 6.41; N, 1.29. Found: C, 68.56; H, 6.30; N, 1.10.

Benzyl O - (Methyl 5 - Acetamido - 4,7,8,9 - tetra - O - acetyl - 3,5 dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2-3)-O-(6-Obenzoyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-O-[2,3,4-tri-O-benzyl- α -L-fucopyranosyl-(1 \rightarrow 4)]-2-acetamido-6-O-benzyl-2-deoxy- β -D-glucopyranoside (36). To a solution of 35 (80.6 mg, 0.08 mmol) and 25 (89 mg, 0.15 mmol) in dry CH₃CN (1 mL) was added MS-4Å (0.1 g), and the mixture was stirred for 8 h at room temperature, then it was cooled to -35 °C. NIS (69 mg, 0.3 mmol) and TfOH (5 μ L, 0.06 mmol) were added to the cooled mixture, and it was stirred for 3 h at -35 °C and then neutralized with Et₃N. A workup similar to that described for 7 and column chromatography (40:1 CH₂Cl₂-MeOH) on silica gel to give 36 (80 mg, 69%) as an amorphous mass: [α]D -41° (c 0.9, CHCl₃); ¹H NMR (CDCl₃) δ 1.18 (d, 3H, J_{5,6} = 6.5 Hz, H-6c), 1.50, 1.88, 2.00 (×2), 2.09, 2.14 (6s, 18H, 2AcN and 4AcO), 1.95 (t, 1H, J = 13 Hz, H-3dax), 2.57 (dd, 1H, J_{gem} = 13, J_{3eq,4} = 4.5 Hz, H-3deq), 3.77 (s, 3H, CO₂Me), and 7.05-7.50 and 8.06 (m+d, 30H, 6Ph). Anal. Calcd for C82H96N2O28 (1557.7): C, 63.23; H, 6.21; N, 1.80. Found: C, 63.06; H, 6.01; N, 1.67.

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REFERENCES AND NOTES

- 1. Synthetic studies on sialoglycoconjugates, Part 93. For Part 92, see H. Ando, H. Ishida, M. Kiso and A. Hasegawa, *Carbohydr. Res.*, in press.
- a) L. A. Lasky, Science, 258, 964 (1992); b) T. Feizi, Curr. Opin. Struct. Biol.,
 3, 701 (1993); c) A. Varki, Proc. Natl. Acad. Sci. U.S.A., 91, 7390 (1994).
 Also see references cited in these articles.
- a) S. Hakomori and R. Kannagi, J. Natl. Cancer Inst., 71, 231 (1983); b) T. Feizi, Nature, 314, 53 (1985); A. Singhal and S. Hakomori, BioEssays, 12, 223 (1990).
- C. -T. Yuen, K. Bezouska, J. O'Brien, M. Stoll, R. Lemonie, A. Lubineau, M. Kiso, A. Hasegawa, N. J. Bockovich, K. C. Nicolaou and T. Feizi, *J. Biol. Chem.*, 269, 1595 (1994).
- a) A. Kameyama, H. Ishida, M. Kiso and A. Hasegawa, Carbohydr. Res., 209, c1 (1991), and J. Carbohydr. Chem., 10, 549 (1991); b) M. Yoshida, A. Uchimura, M. Kiso and A. Hasegawa, Glycoconjugate J., 10, 3 (1993); c) A. Hasegawa, T. Ando, M. Kato, H. Ishida and M. Kiso, Carbohydr. Res., 257, 67 (1994); d) A. Hasegawa and M. Kiso, Methods Enzymol., 242, 158 (1994); e) T. Terada, M. Kiso and A. Hasegawa, Carbohydr. Res., 259, 201 (1994); f) A. Kameyama, T. Ehara, Y. Yamada, H. Ishida, M. Kiso and A. Hasegawa, J. Carbohydr. Chem., 14, 507 (1995); g) S. Komba, H. Ishida, M. Kiso and A. Hasegawa, Glycoconjugate J., 13, 241 (1996).
- a) T. Murase, H. Ishida, M. Kiso and A. Hasegawa, Carbohydr. Res., 184, c1 (1988); b) A. Hasegawa, T. Nagahama, H. Ohki, K. Hotta, H. Ishida and M. Kiso, J. Carbohydr. Chem., 10, 493 (1991); c) A. Hasegawa and M. Kiso, J. Synth. Org. Chem., 50, 429 (1992); d) M. Kiso and A. Hasegawa, Methods Enzymol., 242, 173 (1994); c) A. Hasegawa and M. Kiso in Preparative Carbohydrate Chemistry-A Selection of Themes and Methods: S. Hanessian, Ed.; Marcel Dekker, Inc., New York, 1996, in press.

- a) B.K. Brandley, M. Kiso, S. Abbas, P. Nikrad, O. Srivastava, C. Foxall, Y. Oda and A. Hasegawa, *Glycobiology*, **3**, 633 (1993); b) H. Ohmoto, K. Nakamura, T. Inoue, N. Kondo, Y. Inoue, K. Yoshino, H. Kondo, H. Ishida, M. Kiso and A. Hasegawa, *J. Med. Chem.*, **39**, 1339 (1996); c) A. Hasegawa and M. Kiso in *Carbohydrates: Targets for Drug Design*; Zbigniew J. Witczak and Karl A. Nieforth, Eds.; Marcel Dekker, Inc., New York, 1996, p 138.
- 8. A. Kameyama, H. Ishida, M. Kiso and A. Hasegawa, J. Carbohydr. Chem., 13, 641 (1994).
- 9. M. Kiso, H. Furui, H. Ishida and A. Hasegawa, J. Carbohydr. Chem., 15, 1 (1996).
- 10. K. Toshima and K. Tatsuta, Chem. Rev., 93, 1503 (1993).
- 11. T. Ogawa and S. Nakabayashi, Carbohydr. Res., 97, 81 (1981).
- 12. T.B. Grindley and R. Thangarasa, Can. J. Chem., 68, 1007 (1990).
- a) G. H. Veeneman, S. H. van Leeuwen and J. H. van Boom, *Tetrahedron Lett.*, 31, 1131 (1990); b) P. Konradsson, D. R. Mootoo, R. E. McDevitt and B. Fraser-Reid, J. Chem. Soc. Chem. Commun., 270 (1990).
- 14. R. U. Lemieux and H. Driguez, J. Am. Chem. Soc., 97, 4063 (1975).
- a) A. Marra and P. Sinaÿ, *Carbohydr. Res.*, **187**, 35 (1989); b) A. Hasegawa,
 K. Fushimi, H. Ishida and M. Kiso, *J. Carbohydr. Chem.*, **12**, 1203 (1993); c)
 R. Vig, R. K. Jain, C. F. Piskorz and K. L. Matta, *J. Chem. Soc. Chem. Commun.*, 2073 (1995).
- K. Kitagawa, H. Nakada, Y. Numata, A. Kurosaka, S. Fukui, I. Funakoshi, T. Kawasaki, I. Shimada, F. Inagaki and I. Yamashina, J. Biol. Chem., 265, 4859 (1990).
- a) R. U. Lemieux, D. R. Bundle and D. A. Baker, J. Am. Chem. Soc., 97, 4076 (1975); b) J. Marz and H. Kunz, SYNLETT, 589 (1992); c) A. Lubineau, J. Le Gallic and R. Lemoine, J. Chem. Soc. Chem. Commun., 1419 (1993); d) K. C. Nicolaou, N. J. Bockovich and D. R. Carcanague, J. Am. Chem. Soc., 115, 8843 (1993); e) N. E. Nifant'ev, A. S. Shashkov, Y. E. Tsvetkov, A. B. Tuzikov, I. V. Abramenko, D. F. Gluzman and N. V. Bovin in Synthetic Oligosaccharides-Indispensable Probes for the Life Sciences; P. Kovac, Ed.; ACS Symposium Ser, 560, American Chemical Society, Washington, DC 1994, p 267; f) D. D. Manning, C. R. Bertozzi, S. D. Rosen and L. L. Kiessling, Tetrahedron Lett., 37, 1953 (1996).